

## Botanical Phenolics and Brain Health

Albert Y. Sun · Qun Wang · Agnes Simonyi ·  
Grace Y. Sun

Received: 31 March 2008 / Accepted: 2 October 2008 / Published online: 1 November 2008  
© Humana Press 2008

**Abstract** The high demand for molecular oxygen, the enrichment of polyunsaturated fatty acids in membrane phospholipids, and the relatively low abundance of antioxidant defense enzymes are factors rendering cells in the central nervous system (CNS) particularly vulnerable to oxidative stress. Excess production of reactive oxygen species (ROS) in the brain has been implicated as a common underlying factor for the etiology of a number of neurodegenerative diseases, including Alzheimer's disease (AD), Parkinson's disease (PD), and stroke. While ROS are generated by enzymatic and nonenzymatic reactions in the mitochondria and cytoplasm under normal conditions, excessive production under pathological conditions is associated with activation of  $\text{Ca}^{2+}$ -dependent enzymes including proteases, phospholipases, nucleases, and alterations of signaling pathways which subsequently lead to mitochondrial dysfunction, release of inflammatory factors, and apoptosis. In recent years, there is considerable interest to investigate antioxidative and anti-inflammatory effects of phenolic compounds from different botanical sources. In this review, we describe oxidative mechanisms associated with AD, PD, and stroke, and evaluate neuroprotective effects of phenolic compounds, such as resveratrol from

grape and red wine, curcumin from turmeric, apocynin from *Picrorhiza kurroa*, and epi-gallocatechin from green tea. The main goal is to provide a better understanding of the mode of action of these compounds and assess their use as therapeutics to ameliorate age-related neurodegenerative diseases.

**Keywords** Polyphenols · Neurodegenerative diseases · Oxidative stress · Neuroinflammation · NADPH oxidase · Phospholipase A2 · Mitochondria dysfunction · Alzheimer · Parkinson · Stroke

### Introduction

Reactive oxygen species (ROS) and reactive nitrogen species (RNS), such as superoxide anion, hydroxyl radicals, hydrogen peroxide, lipid peroxyl radicals, nitric oxide, and peroxynitrite, are generated in different cellular systems through enzymatic and nonenzymatic reactions (Sun and Chen 1998). Many pathological conditions are associated with excessive production of ROS/RNS which can attack key proteins, lipids and DNA, alter signal transduction pathways, destroy membranes and subcellular organelles, and subsequently result in apoptosis and cell death. In the presence of transition metals or redox cycling compounds (including quinones), ROS such as superoxide can be converted to the more reactive hydroxy radicals. In some cellular conditions, superoxide anions and nitric oxide can react with each other and form peroxynitrite, a highly toxic anionic compound.

A number of intracellular enzymes are known to produce ROS/RNS, e.g., xanthine/xanthine oxidase, NADPH oxidase, cytochrome P450, nitric oxide synthases, prostaglandin synthases, and enzymes in the electron transport

---

A. Y. Sun · Q. Wang  
Department of Medical Pharmacology and Physiology,  
University of Missouri, Columbia, MO 65211, USA

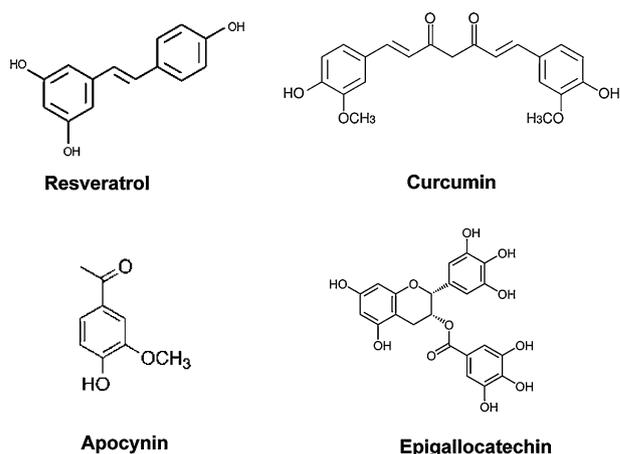
A. Y. Sun · G. Y. Sun  
Department of Pathology and Anatomical Sciences,  
University of Missouri, Columbia, MO 65211, USA

A. Simonyi · G. Y. Sun (✉)  
Department of Biochemistry, University of Missouri,  
117 Schweitzer Hall, Columbia, MO 65211, USA  
e-mail: sung@missouri.edu

chain in mitochondria. In the cellular/subcellular systems, however, production of ROS/RNS through these oxidative enzymes can be counteracted by intracellular antioxidants, including glutathione, vitamin C and E, Coenzyme Q, and by antioxidant enzymes such as superoxide dismutases (SOD), catalase, and glutathione peroxidase. Recent studies also recognize the role of protein kinases and signaling molecules in regulating transcription factors, such as NF $\kappa$ B and Nrf-2/ARE, and thus genes involved in inflammation and oxidant responses (Lim et al. 2007a; Mattson 2008).

The high demand for molecular oxygen, the high levels of polyunsaturated fatty acids in neural membrane phospholipids, and the high iron content are important factors rendering cells in the central nervous system (CNS) to oxidative stress. Oxidative stress is an important underlying factor for a number of neurodegenerative diseases (Halliwell 2006). Neurons are particularly at risk to oxidative stress because many major antioxidant defense mechanisms, such as GSH, Nrf-2, and metallothienin, seem to be localized to astrocytes. Excessive ROS production is associated with activation of the Ca<sup>2+</sup>-dependent enzymes including proteases, phospholipases, and nucleases and alterations of signaling pathways that lead to mitochondrial dysfunction and neuronal apoptosis (Mattson 2007). Increase in oxidative products, such as 4-hydroxynonenal (HNE) for lipid peroxidation, 3-nitrotyrosine (3-NT) for protein carbonyl and protein nitrotyrosine adducts, and 8-hydroxy-deoxyguanosine (8-OHdG) for DNA damage, associated with neurodegenerative diseases support the notion that oxidative stress is a common element in the progression of these diseases (Halliwell 2006; Simonian and Coyle 1996; Sun and Chen 1998).

Oxidative stress is also a significant factor associated with the decline of function in the aging brain. With the disproportional increase in aging population (baby boomers) in the next decade, there is increasing attention to develop nutritional therapies to combat these age-related oxidative processes. Considerable attention is focused on botanicals in vegetables, fruits, grains, roots, flowers, seeds, tea, and red wine. Other nutritional interventions such as dietary restriction and a Mediterranean diet have also captured considerable attention, in particular among older population and subjects with mild cognitive impairments (Burgener et al. 2008). Compounds such as resveratrol from grape and wine, curcumin from turmeric, and epigallocatechin from green tea are becoming recognized for their protective effects against inflammatory diseases, cancers, cardiovascular, and neurodegenerative diseases. Although the mechanisms whereby these compounds display beneficial effects remain elusive, there is increasing evidence to support their antioxidative, anti-inflammatory, antiapoptotic, and metal-chelating properties (Rice-Evans and Miller 1997; Ndiaye et al. 2005). Besides



**Fig. 1** Molecular structure of resveratrol, curcumin, apocynin, and epi-gallocatechin-gallate

these polyphenolic compounds, there is increasing evidence for NADPH oxidase as an important source of ROS in the CNS. Recent studies also place emphasis on ability for apocynin, a phenolic compound derived from *Picrorhiza kurroa*, to inhibit NADPH oxidase (Fig. 1). The major goal for this review is to describe oxidative mechanisms underlying neurodegenerative diseases such as AD, PD, and stroke and to assess whether these phenolic compounds may offer neuroprotective effects.

## Oxidative Stress and Neurodegenerative Disorders

### Alzheimer's Disease

Alzheimer's disease (AD) is the most common form of dementia affecting more than 4 million people in the U.S. and 15–20 million worldwide. With the disproportional increase in the aging population in the next decade, these numbers are projected to triple by 2050. Common pathological hallmarks for AD are accumulation of amyloid plaques and neurofibrillary tangles (McKeel et al. 2004). Besides genetic factors which comprise around 7% of familial AD patients (FAD), epi-genetic and environmental factors are known to play an important role in the onset of sporadic AD. Cardiovascular abnormalities such as hypertension, diabetes, mini-stroke, and atherosclerosis are factors precipitating the increased risk for AD.

Because increase in oxidative stress is associated with early development of AD (Butterfield et al. 2002), there is interest to search for effective therapy to combat the oxidative damage in this disease. There is evidence that at least part of the oxidative mechanism is contributed by the amyloid beta (A $\beta$ ) peptides. These peptides (39–43 amino acids) are released from the amyloid precursor

protein through beta and gamma secretases and upon release, can be aggregated to oligomeric form. Oligomeric Abeta can confer oxidative insult to neurons and glial cells and initiate changes in synaptic plasticity, events occurring long before their deposition to form the amyloid plaques (Selkoe 2001). Although the mechanism for oligomeric Abeta to confer cytotoxicity that results in synaptic dysfunction is not clearly understood, there is evidence that these peptides can confer specific action on the *N*-methyl-D-aspartic acid (NMDA) receptors (Snyder et al. 2005). Aside from regulating synaptic plasticity and memory function, activation of NMDA receptor is coupled to ROS production (Kishida and Klann 2007; Kishida et al. 2005). Recent studies further demonstrate that Abeta can induce ROS production in neurons through an NMDA receptor-dependent process (De Felice et al. 2007; Shelat et al. 2008). Thus, NADPH oxidase may be common in NMDA- and Abeta-induced ROS production, and activation of signaling pathways, including PKC and MAPK, which in turn, lead to activation of cytosolic phospholipase A2 (cPLA<sub>2</sub>) and release of arachidonic acid (AA) (Shelat et al. 2008). Arachidonic acid not only is a precursor for synthesis of prostaglandins, but is also known to serve as a retrograde transmitter in regulating synaptic plasticity (Sang and Chen 2006). Studies by Kriem et al. (2005) demonstrated the involvement of cPLA<sub>2</sub> in Abeta-induced apoptosis in neurons (Kriem et al. 2005).

Intracellular Abeta may target cytoplasmic signaling pathways and impair mitochondrial function (Wang et al. 2007). In astrocytes, Abeta treatment was shown to cause the decrease in mitochondrial membrane potential, and this was partly due to activation of phospholipase A2 (Zhu et al. 2006). In most instances, mitochondrial dysfunction is associated with increase production of ROS and release of cytochrome C, which in turn, triggers the apoptotic pathways. Abeta-mediated ROS production is also linked to increased inflammatory responses, including increased production of cytokines, nitric oxide, and eicosanoids (Mancuso et al. 2007; Butterfield et al. 2002; Akama and Van Eldik 2000). Other contributions from astrocytes include alterations in the synthesis of ApoE (major risk factor for AD) and D-serine, which is an endogenous activator of NMDA receptors.

NADPH oxidase has been regarded as an important source of ROS that mediate the inflammatory responses in astrocytes and microglial cells in the brain. In fact, Abeta-induced ROS from NADPH oxidase in astrocytes is a key factor in mediating neuronal death (Abramov et al. 2004). Therefore, there is strong rationale to develop antioxidant strategy to ameliorate the inflammatory responses associated with the progression of AD. Many recent studies have provided compelling evidence to support dietary supplement of polyphenolic compounds from plant sources to

minimize the oxidative events in the AD brain (Anekonda 2006; Chauhan and Sandoval 2007; Ringman et al. 2005). These herbal alternatives may provide greater therapeutic benefit compared to a single-ingredient synthetic pharmaceutical drug which normally has serious side effects (Kotilinek et al. 2008). Table 1 (top) provides a summary of recent studies testing different botanicals on AD models.

#### Parkinson's Disease

Parkinson's disease (PD) affects approximately 1% of the population over the age of 50. The clinical manifestations of PD include tremors, bradykinesia, muscle rigidity, and akinesia. The pathological landmarks include a progressive loss of dopaminergic neurons in the substantia nigra (Cardoso et al. 2005). Despite numerous hypotheses and speculations for the etiology of PD, oxidative stress remains the strongest leading theory (Miller et al. 2008).

Increased risk for PD is correlated with exposure to environmental factors including heavy metals and herbicides (Brooks et al. 1999; Liou et al. 1997; Yang and Sun 1998b). MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) is an environmental toxin which can selectively damage the substantia nigra and produces Parkinson-like symptoms in animal models and in humans. Studies with this PD model have provided important information about the possible cause of PD (Adams and Odunze 1991; Langston et al. 1987; Schapira 1996). Besides MPTP, other environmental toxins including rotenone, manganese (Sun et al. 1993), dimethoxyphenyl-ethylamine (DMPEA) (Koshimura et al. 1997), and paraquat (Li and Sun 1999; Yang and Sun 1998a, b) also target dopamine neurons. These agents can make their way to the substantia nigra and induce apoptotic pathways in dopaminergic neurons (Schober 2004; Lim et al. 2007b).

Dopamine is a neurotransmitter that can undergo metabolism either by monoamine oxidase (MAO) or by autooxidation, producing H<sub>2</sub>O<sub>2</sub>, superoxide anion, and hydroxyl radicals. In addition, nitric oxide, which is produced through inflammation-induced microglia activation or excitotoxic insults (Abekawa et al. 1997; Gonzalez-Hernandez et al. 1996), may also play a role in the pathogenesis of PD. Formation of peroxynitrite anions through the combination of ROS with nitric oxide may confer additional toxicity to dopaminergic neurons.

Microglia activation is an important factor contributing to the inflammatory responses in PD (Castano et al. 1998; Gao et al. 2003b). Earlier studies demonstrated higher levels of microglia in the PD brain as compared to the age-matched control brain (McGeer et al. 1988). Activated microglia are present in the substantia nigra in several models of PD, including those induced by exposure to MPTP, rotenone, and 6-OHDA (Block et al. 2006; Gao

**Table 1** Effects of common botanicals on AD, PD, and stroke

Polyphenol/plant name	Model	Effects	References
<i>AD models</i>			
Blueberry	Tg2576 mice	+	Joseph et al. (2003)
EGCG	Tg2576 mice	+	Rezai-Zadeh et al. (2005)
Garlic	Tg2576 mice	+	Chauhan (2003, 2006)
	TgCRND8 mice	+	Chauhan and Sandoval (2007)
<i>Ginkgo biloba</i>	Tg2576 mice	+	Stackman et al. (2003)
	TgAPP/PS1 mice	+	Garcia-Alloza et al. (2006) and Tchantchou et al. (2007)
Ginseng	Tg2576 mice	+	Chen et al. (2006)
Ginsenoside Rb1 or M1	A $\beta$ infusion (i.c.v.) mice	+	Tohda et al. (2004)
Pomegranate	Tg2576 mice	+	Hartman et al. (2006)
<i>PD models</i>			
Black tea	6-OHDA rat	+	Chaturvedi et al. (2006)
Cocoa	6-OHDA rat	-/+	Datla et al. (2007)
EGCG	MPTP mice	+	Choi et al. (2002) and Mandel and Youdim (2004)
<i>Ginkgo biloba</i>	6-OHDA rat	+	Kim et al. (2004)
Grape seed	6-OHDA rat	-	Datla et al. (2007)
Green tea	MPTP mice	+	Choi et al. (2002)
	6-OHDA rat	+	Guo et al. (2007)
Quercetin	6-OHDA rat	-	Zbarsky et al. (2005)
Red clover	6-OHDA rat	+	Datla et al. (2007)
Tangerine peel extract	6-OHDA rat	+	Datla et al. (2007)
<i>Stroke models</i>			
Blubberies	Rat permanent left CCAO+H	+	Sweeney et al. (2002)
	Rat transient MCAO	+	Wang et al. (2005d)
Buckwheat polyphenols	Rat repeated ischemia	+	Pu et al. (2004)
Curcuma oil	Rat transient MCAO	+	Rathore et al. (2008)
Garlic	Rat transient MCAO	+	Saleem et al. (2006)
Ginseng	Gerbil transient CCAO	+	Shen and Zhang (2003)
Grape seed extract	Neonatal rat H-I	+	Feng et al. (2005, 2007)
Green tea extract	Rat transient MCAO	+	Hong et al. (2000)
	Gerbil transient CCAO	+	Hong et al. (2001)
Mulberry extract	Mouse transient MCAO	+	Kang et al. (2006)
Pomegranate	Neonatal mouse H-I	+	West et al. (2007)
Red wine polyphenols	Rat transient MCAO	+	Ritz et al. (2008)
Sesame oil	Rat transient MCAO	+	Ahmad et al. (2006)
Spinach	Rat transient MCAO	+	Wang et al. (2005d)

et al. 2002). Abnormal accumulation of iron in microglia and increased levels of  $\alpha$ -synuclein are important pathological features in these models. The ability for microglia to produce high levels of ROS through NADPH oxidase is regarded as an important factor underlying the MPTP-induced dopaminergic neurodegeneration (Gao et al. 2003a, b; Mander et al. 2006; Wu et al. 2003). In our recent study with BV2 microglial cells, paraquat-induced ROS through NADPH oxidase was shown to require protein kinases such as PKCdelta and ERK1/2 (Miller et al. 2007). In microglia-neuron coculture, microglia lacking functional

NADPH oxidase failed to produce neurotoxicity in response to paraquat (Wu et al. 2005). The important role of microglia in pathogenesis of PD can be demonstrated by the ability for minocycline, an antibiotic known to inhibit microglial activation to attenuate the neurotoxicity caused by rotenone (Casarejos et al. 2006).

A number of studies have demonstrated the protective effects of plant phenolics against brain damage in PD. These studies have used either a single compound such as resveratrol, curcumin, EGCG, or a complex mixture of extracts from grape, blueberry, and green tea (Weinreb

et al. 2004; Mercer et al. 2005; Chen et al. 2007; Masuda et al. 2006). Table 1 provides a summary of the studies using different botanicals on PD models. The neuroprotective effects of these phenolic compounds are attributed in part to the free radical scavenging, iron/metal chelating, and their anti-inflammatory properties. There is evidence that these phenolic compounds can target specific signaling pathways and interact with specific proteins, including aggregation of alpha-synuclein (Masuda et al. 2006; Ramassamy 2006; Vafeiadou et al. 2007).

## Stroke

Stroke is the third leading cause of death and the first cause of disability in aging adults. The primary cause of stroke is the interruption of cerebral blood flow either by an arterial or venous obstruction or a cardiac arrest. The pathological manifestations in stroke are diverse, depending on the severity, duration, and localization of the ischemic damage. In the past, many animal models have been developed in which blood flow is focally or globally, permanently or transiently, completely or incompletely interrupted. The most widely established methods of global cerebral ischemia in rodents (rats, mice, or gerbils) are the 2- and 4-vessel occlusion. In gerbils, occlusion of both common carotid arteries (CCA) for 5 min can cause delayed neuronal death (DND) of pyramidal neurons in the hippocampal CA1 area after 4 days (Wang et al. 2002). In addition, the DND is accompanied by increased reactive astrocytes and microglial cells in the injured area (Wang et al. 2002).

Focal cerebral ischemia is usually produced by occlusion of the middle cerebral artery (MCA), either through surgical exposure of the artery after craniotomy or by inserting a suture from the CCA to the MCA to block the blood flow (Chan et al. 1990). Cessation of cerebral blood flow is accompanied by rapid metabolic changes including decrease in ATP production, neuronal membrane depolarization, and release of excitatory neurotransmitters. Despite obvious limitations with each model, the focal ischemic model appears to reflect the most common form of clinical stroke. In focal ischemia followed by reperfusion (I/R), cerebral infarcts with extensive loss of neurons and activation of glial cells are found within 12–24 h after the insult. The penumbral area surrounding the ischemic core is comprised of a large number of reactive astrocytes and microglial cells. Factors causing activation of glial cells in the penumbral area and their role in preventing the spreading of depression and restoring neuronal function remain to be an important area of study.

Oxidative stress has been regarded as a substantial underlying cause of brain damage and neuronal dysfunction after cerebral I/R (Chan 2001). However, the mechanism(s) underlying ROS production and how neurons and glial cells

respond to I/R has not been clearly elucidated. Earlier studies with neurons in culture demonstrated the role of ionotropic glutamate receptors, particularly the NMDA subtype, in triggering massive  $\text{Ca}^{2+}$  influx and in turn, the activation of  $\text{Ca}^{2+}$ -dependent enzymes that trigger mitochondrial dysfunction and apoptotic cell death (Choi 1992). Although mitochondrial dysfunction is known to produce ROS that causes neuronal apoptosis in cerebral ischemia (Chan 2004), recent studies also provided evidence for the involvement of ROS from NADPH oxidase (Wang et al. 2006b; Tang et al. 2007). To combat the deleterious effects of oxidative stress associated with I/R, a number of studies have attempted to upregulate antioxidant enzymes, e.g., superoxide dismutases, catalase, and glutathione peroxidase. Studies with transgenic mice overexpressing SOD1 or GSH-Px-1 have provided support for an important role of these enzymes to remove superoxide and decrease oxidative injury in both global and transient MCAO ischemic models (Saito et al. 2005).

The underlying role of oxidative stress in neuronal damage after I/R also raises attention to possible beneficial effects of polyphenolic compounds from different plant sources (Bravo 1998; Deschamps et al. 2001; Voko et al. 2003; Youdim and Joseph 2001). Studies suggest that some polyphenols can be preventative as well and may thus can act at multiple levels to influence both the early and late phases in the progression of stroke (Curin et al. 2006; Simonyi et al. 2005). Data in Table 1 provide a summary of recent studies testing different botanicals on stroke models.

## Botanical Phenolics and Neurodegenerative Disorders

The use of plant-derived supplements for improving health is gaining popularity because most people consider these natural products to be safer and produce less side effects than synthetic drugs (Raskin et al. 2002). Today, one in three Americans use herbal supplements; consumption is generally greater among woman, patients undergoing surgery, and elderly men (Ang-Lee et al. 2001; Morelli and Naquin 2002). There are more than 50 different plant species and over 8,000 phenolic compounds identified either in single, pure molecular form or in specific proportions of differing plant extracts. Investigating the health benefits of these natural compounds is an enormous challenge to modern medicine.

Polyphenols such as resveratrol were initially identified as the plant's defensive response against stress from ultraviolet radiation, pathogens, and physical damage (Ferguson 2001). For this and other reasons, the polyphenol content in a specific plant source may vary, and differences in procedures for extraction, processing, and storage may also affect purity of the product and inconsistency in the package product.

Polyphenols are divided into different groups depending on the number of phenol rings and the chemical groups attached to the rings. Flavonoids make up the largest and the most important single group of polyphenols and can be divided into subgroups such as flavanols (catechin, epicatechin), flavonols (quercetin, myricetin, kaempferol), flavanones (hesperetin, naringenin), flavones (apigenin, luteolin), isoflavonoids (genistein, daidzein), and anthocyanins (cyaniding, malvidin). The capacity of flavonoids to act as an antioxidant is dependent upon their molecular structure, the position of hydroxyl groups, and other substitutions in the chemical structure of these polyphenols. A number of excellent reviews dealing with their structure, absorption, metabolism, and pharmacokinetics have been published (Bravo 1998; Ross and Kasum 2002; Manach and Donovan 2004). Besides scavenging free radicals, many phenolics also exhibit multiple biological properties, e.g., anti-inflammatory, anticancer, antiviral, antimicrobial, vasorelaxant, and anticlotting activities (Rahman et al. 2007). In general, these phenolic compounds are rapidly converted to their glucuronide derivatives upon ingestion and are transported to the circulatory system and different body organs including the brain. In recent years, a number of reviews have reported on neuroprotective effects of polyphenols in cell and animal models (Wang et al. 2001; Dajas et al. 2003; Mandel and Youdim 2004; Simonyi et al. 2005). This review is limited to neuroprotective effects of resveratrol from grape and wine, curcumin from turmeric, apocynin from *Picrorhiza kurroa*, and epigallocatechin-3-gallate from green tea (Fig. 1).

There is evidence that some phenolic compounds exert their mode of action and target different intracellular pathways on a concentration-dependent manner. For example, low dose of red wine polyphenols was shown to promote angiogenesis via activation of the Akt/PI3K/eNOS, p38MAPK pathway but not the NF- $\kappa$ B pathway. However, at high dose, they can be antiangiogenic through inhibition of the Akt/PI3K/eNOS pathway and enhancing the NF- $\kappa$ B pathway (Baron-Menguy et al. 2007). Another example is epicatechin, which not only exerts antioxidant activity but also can modulate protein kinase signaling pathways, depending on the concentration of the compound administered. In the study by Schroeter et al. (2007), epicatechin stimulated ERK- and PI3K-dependent CREB phosphorylation at lower concentration of 100–300 nmol/l but this effect was no longer apparent at the higher concentration of 30  $\mu$ mol/l. These dose effects may be important to explain the anti- versus pro-oxidant actions of the phenolics and differences in experimental outcomes from different laboratories. It is also important to recognize that results from studies of phenolic compounds in cell culture system may not correspond to their action in vivo (Halliwell 2008).

## Resveratrol

Epidemiological studies have reported that despite consuming a fatty diet, the population with moderate wine consumption has a lower incidence of cardiovascular diseases. A widely held theory for the cardioprotective effects of the “French paradox” is the antiplatelet aggregation properties of compounds in red wine in preventing the development of atherosclerotic plaques. In recent years, studies further indicated that red wine and grape polyphenols may also offer protective effects against neurodegenerative diseases (Esposito et al. 2002; Simonyi et al. 2002; Sun et al. 1999a, b). Studies from our laboratory provided evidence that dietary supplement of polyphenols extracted from grape skin and seeds could ameliorate oxidative damage in synaptic membranes in the brain induced by chronic alcohol consumption (Sun et al. 1999a, b). Grape polyphenols also prevented chronic ethanol-induced increase in COX-2 mRNA expression in the rat brain (Simonyi et al. 2002).

Although grape also contains other types of polyphenols, trans-resveratrol (3,4',5-trihydroxystilbene) is considered the most effective compound in producing beneficial health effects. In addition to grapes, resveratrol is found in a variety of plant species including peanuts and berries (Baur and Sinclair 2006). Resveratrol is also highly concentrated in some oriental herbal plants, such as *kojo-kan*, *polygonum caspidatum*, which is used to treat fevers, hyperlipidemia, atherosclerosis, and inflammation (Chung et al. 1992). In our studies with PC-12 cells, resveratrol was more effective in protecting against oxidative damage than vitamins E and C combined (Chanvitayapongs et al. 1997). A number of studies using cell models have provided information for the underlying mechanisms for neuroprotective effects of resveratrol (Gao et al. 2006a, b; Lu et al. 2006; Raval et al. 2006; Cho et al. 2008; Tsai et al. 2007). Studies with cell culture models of Parkinson's disease also demonstrated neuroprotective effects of resveratrol in alleviating oxidative damage induced by neurotoxins (Gelinis and Martinoli 2002; Alvira et al. 2007).

Studies from our laboratory demonstrated the ability for resveratrol to protect against ischemia-induced DND in the gerbil global ischemia model (Wang et al. 2002) and neuronal excitotoxicity in rats induced by kainic acid (Wang et al. 2005c). The neuroprotective effects of resveratrol can be demonstrated by different mode of administration, e.g., by i.p. injection and by supplementing as grape powder formulation (Wang et al. 2005a).

Studies to examine bioavailability of resveratrol indicated that this compound is rapidly conjugated to its glucuronide derivative which is probably the vehicle for transportation to the circulatory system. Apparently, this form of resveratrol can readily cross the blood–brain

barrier albeit at lower levels when compared to that in the liver (Wang et al. 2002).

Besides excellent free radical scavenging properties, resveratrol can offer other effects to the cell, e.g., increasing the lifespan in yeast (Howitz et al. 2003). This effect is explained by its ability to activate sirtuins, which belong to a conserved family of NAD<sup>+</sup>-dependent deacetylases (class III histone deacetylases) (Baur and Sinclair 2006). In the lower organisms including yeast, *C. elegans*, and flies, increase in sirtuins is associated with extended lifespan. The multiple roles of resveratrol as an antioxidant and as a life-promoting agent make it an attractive candidate for treatment of neurodegenerative diseases (Anekonda 2006; Mancuso et al. 2007; Baur and Sinclair 2006).

Several studies demonstrated the ability for resveratrol to protect neurons against Abeta-induced toxicity in vitro (Chen et al. 2005; Han et al. 2004; Jang and Surh 2003). In fact, resveratrol combined with other polyphenolic compounds, such as catechin from green tea, can produce synergism in the protective effects (Conte et al. 2003a, b). In a rat model of sporadic AD, chronic administration of resveratrol ameliorated the cognitive impairment and oxidative damage induced by intracerebroventricular injection of streptozotocin (Sharma and Gupta 2002). Red wine consumption also significantly attenuated AD-type deterioration of spatial memory function and Abeta neuropathology in Tg2576 mice (Wang et al. 2006a). There is evidence that resveratrol can inhibit formation and extension of Abeta fibrils and destabilize the fibrilized Abeta (Ono et al. 2006; Ono and Yamada 2006). Another study demonstrated its ability to reduce Abeta secretion in several cell lines via a mechanism that involves the proteasome (Marambaud et al. 2005).

A number of studies have demonstrated the ability of resveratrol to suppress neuroinflammatory responses, e.g., attenuating iNOS and COX-2 expression (Bi et al. 2005; Kim et al. 2006, 2007). However, it is not clear whether this action is related to the ability of resveratrol to minimize ROS production from NADPH oxidase. Consequently, despite strong evidence for therapeutic potential of resveratrol for treatment of cancer, angiogenesis, myocardial infarction as well as different neurodegenerative diseases (Baur and Sinclair 2006), more investigations are needed to understand proper usage of this polyphenol and its mechanism of action on different cell types.

## Curcumin

Curcumin (diferuloylmethane) is derived from turmeric, the powdered rhizome of the medicinal plant *Curcuma longa* Linn. It has been used for centuries throughout Asia as a food additive and a traditional herbal medicine. Recent

studies demonstrated that besides potent antioxidative and anti-inflammatory properties of curcumin, it also exhibits anti-amyloidogenic effects (Ono et al. 2004). Curcumin can bind amyloid directly and inhibit Abeta aggregation as well as prevent fibril and oligomer formation (Yang et al. 2005). These antifibril effects of curcumin were also evidenced in studies with alpha synuclein, the protein involved in PD (Ono and Yamada 2006).

Curcumin supplementation has been recently considered as an alternative, nutritional approach to reduce oxidative, inflammatory damage, and amyloid pathology associated with AD (Wu et al. 2006). However, because curcumin is common in many curry spices and is widely consumed by different populations, it is difficult for well-designed studies to evaluate health effects of this polyphenol. When conventional NSAID, ibuprofen, and curcumin were compared for their ability to protect against Abeta-induced damage, dietary curcumin, not ibuprofen, was shown to suppress oxidative damage and reduced synaptophysin loss (Frautschy et al. 2001). Dietary curcumin also prevented Abeta-induced spatial memory deficits in the Morris water maze and postsynaptic density loss and reduced Abeta deposits (Frautschy et al. 2001). To evaluate whether curcumin could affect Alzheimer-like pathology in Tg2576 mice, both low and high doses of curcumin significantly lowered oxidized proteins and interleukin-1 $\beta$ , a proinflammatory cytokine elevated in the brains of these mice (Lim et al. 2001). Besides its anti-amyloid properties, curcumin can also offer antioxidant, anti-inflammatory, and cholesterol lowering properties, all are important on ameliorating the deleterious consequences of AD (Ringman et al. 2005). Several clinical trials are in progress to address safety, tolerability, and bioavailability of this compound (Ringman et al. 2005; Fiala et al. 2007).

Besides AD, there is in vitro and in vivo data suggesting that curcumin exerts a protective effect against neurodegeneration in cerebral ischemia and Parkinson's disease. In a study in which curcumin was administered through i.v. injection (1 and 2 mg/kg) after focal ischemia, the neuroprotective effects were attributed to a protection of blood-brain barrier integrity (Jiang et al. 2007). In our laboratory, curcumin administered either through i.p. injection (30 mg/kg) or through a dietary supplementation (2.0 g/kg diet) for 2 months significantly attenuated ischemia-induced DND as well as glial cell activation in the gerbil model (Wang et al. 2005b). Most interestingly, curcumin administration not only reduced ischemia-induced lipid peroxidation and mitochondrial dysfunction, it also ameliorated the increase in locomotor activity observed at 48 h after ischemic insult, thus correlating behavioral deficits with the extent of neuronal damage (Wang et al. 2005b). Consistent with other studies, bioavailability study indicated a rapid increase in curcumin in plasma and other body organs including the

brain within 1 h after i.p. injection (Ringman et al. 2005; Goel et al. 2008).

The neuroprotective effects of curcumin relevant to PD are likely to be associated with its antioxidant and anti-inflammatory properties (Chen and Le 2006; Jagatha et al. 2008; Zbarsky et al. 2005). As found for Abeta, curcumin also can inhibit aggregation of alpha-synuclein (Pandey et al. 2008). Recent studies have identified other molecular targets of curcumin, including its action on transcription factors, growth factors, antioxidant enzymes, cell-survival kinases, and signaling molecules (Ramassamy 2006; Salvioli et al. 2007; Goel et al. 2008). On the other hand, it is worth noting that excessive application of curcumin may produce pro-oxidative effects (Ahsan et al. 1999). Therefore, more studies are needed to understand the different modes of action of curcumin on specific enzymes and pathways prior to recommendation for its use as a therapeutic agent.

### Apocynin

Apocynin (4-hydroxy-3-methoxy-acetophenone) was discovered during activity-guided isolation of immunomodulatory constituents from *Picrorhiza kurroa*, a creeping plant native to the mountains of India, Nepal, Tibet, and Pakistan (*Picrorhiza kurroa*, Monograph, 2001). *Picrorhiza kurroa* has been used as an herbal medicine for centuries for treatment of a number of inflammatory diseases. Apocynin may also be obtained from other sources, e.g., from the rhizome of Canadian hemp (*Apocynum cannabinum*), other *Apocynum* species (e.g., *A. androsaemifolium*) or from the rhizomes of *Iris* species. This compound has been regarded as a powerful antioxidant and anti-inflammatory agent, specifically, for blocking the activity of NADPH oxidase through interfering with the assembly of the cytosolic NADPH oxidase components with its membrane components (Stolk et al. 1994).

NADPH oxidase is increasingly recognized for its dual-edge roles in health and disease and has been implicated in the pathogenesis of many diseases, including cardiovascular and neurodegenerative diseases (Bedard and Krause 2007). In recent years, it has become apparent that brain cells constitutively express a superoxide-generating enzyme analogous to the NADPH oxidase in phagocytes (Infanger et al. 2006). The prototypic NADPH oxidase comprises a membrane-associated cytochrome b558 with one p22 phox and one gp91 phox subunit and several regulatory cytosolic subunits (p47 phox, p40 phox, p67 phox, and the small G protein Rac1 or Rac2). Upon phosphorylation, the cytosolic subunits are translocated to bind with the membrane subunits. Consequently, a number of receptor-signaling pathways are linked to activation of

NADPH oxidase leading to rapid production of superoxide anions (Bedard and Krause 2007).

Altered NADPH oxidase function has been linked to neurological disorders such as stroke, Alzheimer's, and Parkinson's diseases (Lambeth 2007). Several reports of human studies (on AD, PD, and stroke) demonstrated upregulation of different subunits expression in microglial cells (Wu et al. 2003). Genetic deletion of gp91phox mitigates neuronal loss in a variety of animal models of neurodegeneration, including the MPTP model of PD and cerebral ischemia (Zhang et al. 2004). Apocynin has been effective in ameliorating neuropathological damages in both in vivo and in vitro models of PD (Anantharam et al. 2007; Gao et al. 2003a, b, c). Apocynin also retarded disease progression and extended survival in a mouse ALS model (Boillee and Cleveland 2008). Immunohistochemical studies demonstrated that the increase in NADPH oxidase subunits expression after transient focal cerebral ischemia is mainly derived from activated microglial cells. Apocynin was effective in preventing ischemic damage and blood–brain barrier disruption in different animal models of experimental stroke (Wang et al. 2006b; Tang et al. 2007; Kahles et al. 2007). In our study using the gerbil global cerebral ischemia model, apocynin inhibited ischemia/reperfusion-induced increase in lipid peroxidation, oxidative DNA damage, and glial cell activation in the hippocampus (Wang et al. 2006b).

NADPH oxidase-dependent production of superoxide radicals has been identified as a major contributor to oxidative and inflammatory responses in the brain under different injury conditions. Activation of NADPH oxidase in glial cells is linked to increased secretion of cytokines and other inflammatory factors (Dringen 2005). Superoxide produced from NADPH oxidase may interact with nitric oxide from iNOS to form the toxic peroxynitrite, which is considered an important factor associated with neuronal death (Brown 2007). Aside from suppressing NF- $\kappa$ B pathway and preventing COX-2 expression in activated monocytes (Barbieri et al. 2004), apocynin is also effective against Abeta-induced microglial proliferation and lipopolysaccharide (LPS) and interferon  $\gamma$ -induced neuronal death (Li et al. 2004; Jekabson et al. 2006; Shibata et al. 2006).

An apparent limitation for therapeutic use of apocynin is the high concentrations needed for exerting beneficial effects. Furthermore, most studies have used acute treatment and few studies employed a preventative, dietary approach. In vitro studies suggest that apocynin may be converted to diapocynin through chemical catalysis using ferrous sulfate and sodium persulfate or through peroxidases such as myeloperoxidase. However, our recent study failed to detect diapocynin in rat plasma and tissues after systemic injection of apocynin (Wang et al. 2008). However, our study on bioavailability showed that similar to other polyphenols,

apocynin is rapidly converted to its glucuronide derivative and transported to the circulation system and other body organs, including the brain. More studies are necessary for considering the potential therapeutic use of apocynin for treatment of neurodegenerative disorders.

Other Natural Phenolics

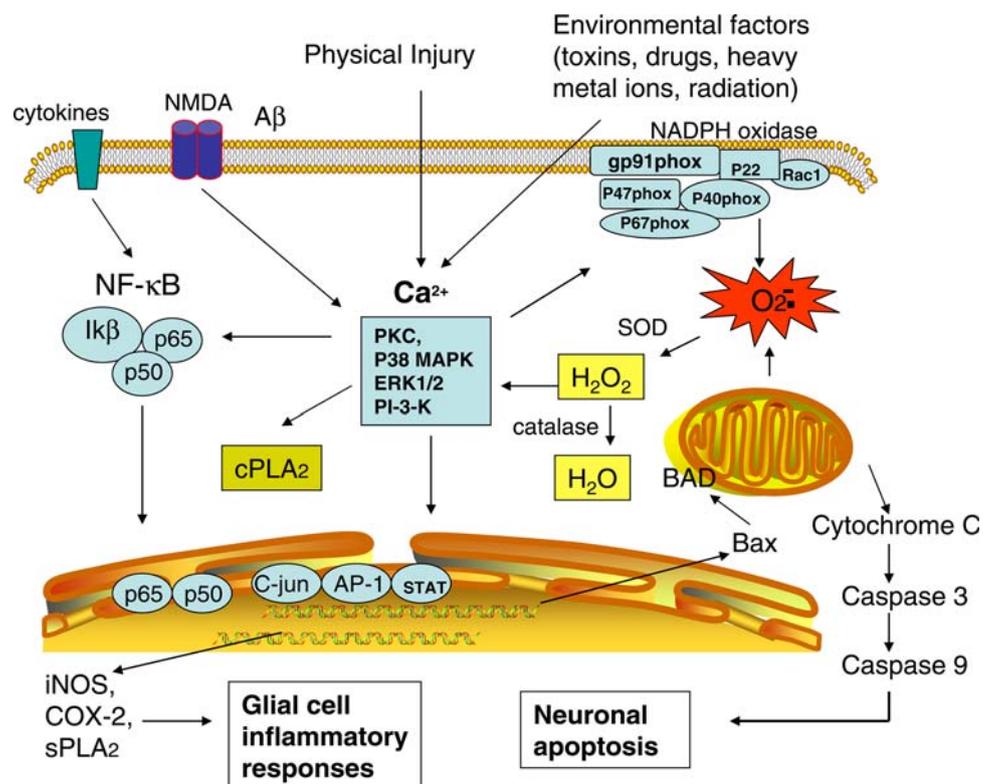
Many other phenolic compounds in fruits and vegetables are good candidates for consideration as therapeutics in combating aging and neurodegenerative diseases (Vauzour et al. 2007). Among these, there is special interest regarding the neuroprotective actions of (–)-epigallocatechin-3-gallate (EGCG) from green tea (Sutherland et al. 2006). Besides its free radical scavenging, iron chelating, and anti-inflammatory properties, EGCG can exert its action on different sites of the apoptotic pathways, including altering the expression of anti- and pro-apoptotic genes. These studies further implicate that green tea extract may also exert protection through controlling calcium homeostasis, activation of MAPK, PKC, antioxidant enzymes, survival genes, and modulating enzymes for processing of the amyloid precursor protein (Mandel et al. 2004; Mandel and Youdim 2004; Weinreb et al. 2008). EGCG was shown to inhibit 6-OHDA-induced NF-κB-mediated expression of cell death and cell cycle genes (Levites et al. 2002a, b).

In light of the neuroprotective effects from different polyphenols and plant extracts, a summary of recent studies describing neuroprotective effects of different botanical compounds in different animal models for AD, PD, and stroke is provided in Table 1. Amentoflavonoid, a naturally occurring bioflavonoid, was able to rescue neurons from hypoxic-ischemic injury (Shin et al. 2006; Yi et al. 2006). This compound seems to implement multiple mechanisms including direct blockade of cell death cascades and anti-inflammatory inhibition of microglia. Coenzyme Q (CoQ) is enriched in a number of diets and is a potent antioxidant. This redox active compound has been implicated to play an important role in improving mitochondrial function. However, whether CoQ(10) can be used as a therapeutic agent for treatment of PD remains to be investigated (Storch et al. 2007).

Botanical Phenolics on Intracellular Signaling Pathways

It is becoming recognized that besides their antioxidative and anti-inflammatory properties, many phenolics may also have specific action on intracellular signaling pathways (Fig. 2). These signaling pathways are interrelated and are evolved from ROS from NADPH oxidase and mitochondria. In particular, these signaling pathways are

Fig. 2 Signaling pathways associated with oxidative stress



downstream of ROS produced from NADPH oxidase upon injury due to cerebral ischemia, Abeta, and excitotoxicity. In our studies, we further link these kinases to activation of cPLA<sub>2</sub> and release of arachidonic acid. There is also evidence that ROS produced from NADPH oxidase is linked to transcriptional pathways, such as the NF- $\kappa$ B pathway and the Nrf/ARE pathway for induction of antioxidant and inflammatory genes (Santangelo et al. 2007) and subsequently, triggering the apoptotic pathway (Zhu et al. 2006). Successful identification of these compounds and their action on intracellular signaling pathways will be important for effective use to combat neurodegenerative diseases.

### Concluding Remarks

Despite complex and diverse genetic and epi-genetic factors underlying manifestations of different neurodegenerative diseases, there are strong reasons to believe that oxidative stress is a common factor playing a central role in the pathogenesis of these diseases. While many pathological conditions are associated to ROS production from mitochondria, more recent studies have unveiled an important role of ROS from NADPH oxidase. Studies here indicate that phenolic compounds such as resveratrol from grape and wine, curcumin from turmeric, epi-gallocatechin from green tea, and apocynin from *Picrorhiza kurroa*, not only exhibit potent antioxidative properties for scavenging free radicals, but may also act on specific signaling pathways for regulating inflammatory responses. These studies support the use of plant-derived phenolic supplements in promoting general health and preventing age-related diseases in humans.

**Acknowledgment** This work was supported by grants (P02 AG018357 and 1R21AT003859) from NIH.

### References

- Abekawa, T., Ohmori, T., & Koyama, T. (1997). Effect of no synthesis inhibition on striatal dopamine release and stereotyped behavior induced by a single administration of methamphetamine. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 21, 831–838. doi:10.1016/S0278-5846(97)00083-3.
- Abramov, A. Y., Canevari, L., & Duchen, M. R. (2004). Beta-amyloid peptides induce mitochondrial dysfunction and oxidative stress in astrocytes and death of neurons through activation of NADPH oxidase. *Journal of Neuroscience*, 24, 565–575. doi:10.1523/JNEUROSCI.4042-03.2004.
- Adams, J. D., Jr., & Odunze, I. N. (1991). Biochemical mechanisms of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine toxicity. Could oxidative stress be involved in the brain? *Biochemical Pharmacology*, 41, 1099–1105. doi:10.1016/0006-2952(91)90646-M.
- Ahmad, S., Yousuf, S., Ishrat, T., Khan, M. B., Bhatia, K., Fazli, I. S., et al. (2006). Effect of dietary sesame oil as antioxidant on brain hippocampus of rat in focal cerebral ischemia. *Life Sciences*, 79, 1921–1928. doi:10.1016/j.lfs.2006.06.017.
- Ahsan, H., Parveen, N., Khan, N. U., & Hadi, S. M. (1999). Pro-oxidant, anti-oxidant and cleavage activities on DNA of curcumin and its derivatives demethoxycurcumin and bisdemethoxycurcumin. *Chemico-Biological Interactions*, 121, 161–175. doi:10.1016/S0009-2797(99)00096-4.
- Akama, K. T., & Van Eldik, L. J. (2000). Beta-amyloid stimulation of inducible nitric-oxide synthase in astrocytes is interleukin-1beta- and tumor necrosis factor-alpha (TNFalpha)-dependent, and involves a TNFalpha receptor-associated factor- and NFkappaB-inducing kinase-dependent signaling mechanism. *Journal of Biological Chemistry*, 275, 7918–7924. doi:10.1074/jbc.275.11.7918.
- Alvira, D., Yeste-Velasco, M., Folch, J., Verdaguier, E., Canudas, A. M., Pallas, M., et al. (2007). Comparative analysis of the effects of resveratrol in two apoptotic models: Inhibition of complex I and potassium deprivation in cerebellar neurons. *Neuroscience*, 147, 746–756. doi:10.1016/j.neuroscience.2007.04.029.
- Anantharam, V., Kaul, S., Song, C., Kanthasamy, A., & Kanthasamy, A. G. (2007). Pharmacological inhibition of neuronal NADPH oxidase protects against 1-methyl-4-phenylpyridinium (MPP+)-induced oxidative stress and apoptosis in mesencephalic dopaminergic neuronal cells. *Neurotoxicology*, 28, 988–997. doi:10.1016/j.jneuro.2007.08.008.
- Anekonda, T. S. (2006). Resveratrol—A boon for treating Alzheimer's disease? *Brain Research Reviews*, 52, 316–326. doi:10.1016/j.brainresrev.2006.04.004.
- Ang-Lee, M. K., Moss, J., & Yuan, C. S. (2001). Herbal medicines and perioperative care. *JAMA*, 286, 208–216. doi:10.1001/jama.286.2.208.
- Barbieri, S. S., Cavalca, V., Eligini, S., Brambilla, M., Caiani, A., Tremoli, E., et al. (2004). Apocynin prevents cyclooxygenase 2 expression in human monocytes through NADPH oxidase and glutathione redox-dependent mechanisms. *Free Radical Biology and Medicine*, 37, 156–165. doi:10.1016/j.freeradbiomed.2004.04.020.
- Baron-Menguy, C., Bocquet, A., Guihot, A. L., Chappard, D., Amiot, M. J., Andriantsitohaina, R., et al. (2007). Effects of red wine polyphenols on postischemic neovascularization model in rats: Low doses are proangiogenic, high doses anti-angiogenic. *FASEB Journal*, 21, 3511–3521. doi:10.1096/fj.06-7782.com.
- Baur, J. A., & Sinclair, D. A. (2006). Therapeutic potential of resveratrol: The in vivo evidence. *Nature Reviews. Drug Discovery*, 5, 493–506. doi:10.1038/nrd2060.
- Bedard, K., & Krause, K. H. (2007). The NOX family of ROS-generating NADPH oxidases: Physiology and pathophysiology. *Physiological Reviews*, 87, 245–313. doi:10.1152/physrev.00044.2005.
- Bi, X. L., Yang, J. Y., Dong, Y. X., Wang, J. M., Cui, Y. H., Ikeshima, T., et al. (2005). Resveratrol inhibits nitric oxide and TNF-alpha production by lipopolysaccharide-activated microglia. *International Immunopharmacology*, 5, 185–193. doi:10.1016/j.intimp.2004.08.008.
- Block, M. L., Li, G., Qin, L., Wu, X., Pei, Z., Wang, T., et al. (2006). Potent regulation of microglia-derived oxidative stress and dopaminergic neuron survival: Substance P vs. dynorphin. *FASEB Journal*, 20, 251–258. doi:10.1096/fj.05-4553.com.
- Boillee, S., & Cleveland, D. W. (2008). Revisiting oxidative damage in ALS: Microglia, Nox, and mutant SOD1. *Journal of Clinical Investigation*, 118, 474–478.
- Bravo, L. (1998). Polyphenols: Chemistry, dietary sources, metabolism, and nutritional significance. *Nutrition Reviews*, 56, 317–333.
- Brooks, A. I., Chadwick, C. A., Gelbard, H. A., Cory-Slechta, D. A., & Federoff, H. J. (1999). Paraquat elicited neurobehavioral syndrome caused by dopaminergic neuron loss. *Brain Research*, 823, 1–10. doi:10.1016/S0006-8993(98)01192-5.

- Brown, G. C. (2007). Mechanisms of inflammatory neurodegeneration: iNOS and NADPH oxidase. *Biochemical Society Transactions*, 35, 1119–1121. doi:10.1042/BST0351166.
- Burgener, S. C., Buettner, L., Coen Buckwalter, K., et al. (2008). Evidence supporting nutritional interventions for persons in early stage Alzheimer's disease (AD). *The Journal of Nutrition, Health & Aging*, 12, 18–21.
- Butterfield, D. A., Griffin, S., Munch, G., & Pasinetti, G. M. (2002). Amyloid beta-peptide and amyloid pathology are central to the oxidative stress and inflammatory cascades under which Alzheimer's disease brain exists. *Journal of Alzheimer's Disease*, 4, 193–201.
- Cardoso, S. M., Moreira, P. I., Agostinho, P., Pereira, C., & Oliveira, C. R. (2005). Neurodegenerative pathways in Parkinson's disease: Therapeutic strategies. *Current Drug Targets. CNS and Neurological Disorders*, 4, 405–419. doi:10.2174/1568007054546072.
- Casarejos, M. J., Menendez, J., Solano, R. M., Rodriguez-Navarro, J. A., Garcia de Yebenes, J., & Mena, M. A. (2006). Susceptibility to rotenone is increased in neurons from parkin null mice and is reduced by minocycline. *Journal of Neurochemistry*, 97, 934–946. doi:10.1111/j.1471-4159.2006.03777.x.
- Castano, A., Herrera, A. J., Cano, J., & Machado, A. (1998). Lipopolysaccharide intranigral injection induces inflammatory reaction and damage in nigrostriatal dopaminergic system. *Journal of Neurochemistry*, 70, 1584–1592.
- Chan, P. H. (2001). Reactive oxygen radicals in signaling and damage in the ischemic brain. *Journal of Cerebral Blood Flow and Metabolism*, 21, 2–14. doi:10.1097/00004647-200101000-00002.
- Chan, P. H. (2004). Mitochondria and neuronal death/survival signaling pathways in cerebral ischemia. *Neurochemical Research*, 29, 1943–1949. doi:10.1007/s11064-004-6869-x.
- Chan, P. H., Fishman, R. A., Wesley, M. A., & Longar, S. (1990). Pathogenesis of vasogenic edema in focal cerebral ischemia. Role of superoxide radicals. *Advances in Neurology*, 52, 177–183.
- Chanvitayapongs, S., Draczynska-Lusiak, B., & Sun, A. Y. (1997). Amelioration of oxidative stress by antioxidants and resveratrol in PC12 cells. *NeuroReport*, 8, 1499–1502. doi:10.1097/00001756-199704140-00035.
- Chaturvedi, R. K., Shukla, S., Seth, K., Chauhan, S., Sinha, C., Shukla, Y., et al. (2006). Neuroprotective and neurorescue effect of black tea extract in 6-hydroxydopamine-lesioned rat model of Parkinson's disease. *Neurobiology of Disease*, 22, 421–434. doi:10.1016/j.nbd.2005.12.008.
- Chauhan, N. B. (2003). Anti-amyloidogenic effect of *Allium sativum* in Alzheimer's transgenic model Tg2576. *Journal of Herbal Pharmacotherapy*, 3, 95–107. doi:10.1300/J157v03n01\_05.
- Chauhan, N. B. (2006). Effect of aged garlic extract on APP processing and tau phosphorylation in Alzheimer's transgenic model Tg2576. *Journal of Ethnopharmacology*, 108, 385–394. doi:10.1016/j.jep.2006.05.030.
- Chauhan, N. B., & Sandoval, J. (2007). Amelioration of early cognitive deficits by aged garlic extract in Alzheimer's transgenic mice. *Phytotherapy Research*, 21, 629–640. doi:10.1002/ptr.2122.
- Chen, F., Eckman, E. A., & Eckman, C. B. (2006). Reductions in levels of the Alzheimer's amyloid beta peptide after oral administration of ginsenosides. *FASEB Journal*, 20, 1269–1271. doi:10.1096/fj.05-5530fje.
- Chen, H., Zhang, M., Qu, Z., & Xie, B. (2007). Compositional analysis and preliminary toxicological evaluation of a tea polysaccharide conjugate. *Journal of Agricultural and Food Chemistry*, 55, 2256–2260. doi:10.1021/jf0632740.
- Chen, J., Zhou, Y., Mueller-Steiner, S., Chen, L. F., Kwon, H., Yi, S., et al. (2005). SIRT1 protects against microglia-dependent amyloid-beta toxicity through inhibiting NF-kappaB signaling. *Journal of Biological Chemistry*, 280, 40364–40374. doi:10.1074/jbc.M509329200.
- Chen, S., & Le, W. (2006). Neuroprotective therapy in Parkinson disease. *American Journal of Therapeutics*, 13, 445–457. doi:10.1097/01.mjt.0000174353.28012.a7.
- Cho, I. J., Ahn, J. Y., Kim, S., Choi, M. S., & Ha, T. Y. (2008). Resveratrol attenuates the expression of HMG-CoA reductase mRNA in hamsters. *Biochemical and Biophysical Research Communications*, 367, 190–194. doi:10.1016/j.bbrc.2007.12.140.
- Choi, D. W. (1992). Excitotoxic cell death. *Journal of Neurobiology*, 23, 1261–1276. doi:10.1002/neu.480230915.
- Choi, J. Y., Park, C. S., Kim, D. J., Cho, M. H., Jin, B. K., Pie, J. E., et al. (2002). Prevention of nitric oxide-mediated 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced Parkinson's disease in mice by tea phenolic epigallocatechin 3-gallate. *Neurotoxicology*, 23, 367–374. doi:10.1016/S0161-813X(02)00079-7.
- Chung, M. I., Teng, C. M., Cheng, K. L., Ko, F. N., & Lin, C. N. (1992). An antiplatelet principle of *Veratrum formosanum*. *Planta Medica*, 58, 274–276. doi:10.1055/s-2006-961453.
- Conte, A., Pellegrini, S., & Tagliazucchi, D. (2003a). Effect of resveratrol and catechin on PC12 tyrosine kinase activities and their synergistic protection from beta-amyloid toxicity. *Drugs Under Experimental and Clinical Research*, 29, 243–255.
- Conte, A., Pellegrini, S., & Tagliazucchi, D. (2003b). Synergistic protection of PC12 cells from beta-amyloid toxicity by resveratrol and catechin. *Brain Research Bulletin*, 62, 29–38. doi:10.1016/j.brainresbull.2003.08.001.
- Curin, Y., Ritz, M. F., & Andriantsitohaina, R. (2006). Cellular mechanisms of the protective effect of polyphenols on the neurovascular unit in strokes. *Cardiovascular & Hematological Agents in Medicinal Chemistry*, 4, 277–288. doi:10.2174/187152506778520691.
- Dajas, F., Rivera, F., Blasina, F., Arredondo, F., Echeverry, C., Lafon, L., et al. (2003). Cell culture protection and in vivo neuroprotective capacity of flavonoids. *Neurotoxicity Research*, 5, 425–432.
- Datla, K. P., Zbarsky, V., Rai, D., Parkar, S., Osakabe, N., Aruoma, O. I., et al. (2007). Short-term supplementation with plant extracts rich in flavonoids protect nigrostriatal dopaminergic neurons in a rat model of Parkinson's disease. *Journal of the American College of Nutrition*, 26, 341–349.
- De Felice, F. G., Velasco, P. T., Lambert, M. P., Viola, K., Fernandez, S. J., Ferreira, S. T., et al. (2007). Abeta oligomers induce neuronal oxidative stress through an N-methyl-D-aspartate receptor-dependent mechanism that is blocked by the Alzheimer drug memantine. *Journal of Biological Chemistry*, 282, 11590–11601. doi:10.1074/jbc.M607483200.
- Deschamps, V., Barberger-Gateau, P., Peuchant, E., & Orgogozo, J. M. (2001). Nutritional factors in cerebral aging and dementia: Epidemiological arguments for a role of oxidative stress. *Neuroepidemiology*, 20, 7–15. doi:10.1159/000054752.
- Dringen, R. (2005). Oxidative and antioxidative potential of brain microglial cells. *Antioxidants & Redox Signaling*, 7, 1223–1233. doi:10.1089/ars.2005.7.1223.
- Esposito, E., Rotilio, D., Di Matteo, V., Di Giulio, C., Cacchio, M., & Algeri, S. (2002). A review of specific dietary antioxidants and the effects on biochemical mechanisms related to neurodegenerative processes. *Neurobiology of Aging*, 23, 719–735. doi:10.1016/S0197-4580(02)00078-7.
- Feng, Y., Liu, Y. M., Fratkins, J. D., & LeBlanc, M. H. (2005). Grape seed extract suppresses lipid peroxidation and reduces hypoxic ischemic brain injury in neonatal rats. *Brain Research Bulletin*, 66, 120–127. doi:10.1016/j.brainresbull.2005.04.006.
- Feng, Y., Liu, Y. M., Leblanc, M. H., Bhatt, A. J., & Rhodes, P. G. (2007). Grape seed extract given three hours after injury

- suppresses lipid peroxidation and reduces hypoxic-ischemic brain injury in neonatal rats. *Pediatric Research*, 61, 295–300. doi:10.1203/pdr.0b013e318030c92d.
- Ferguson, L. R. (2001). Role of plant polyphenols in genomic stability. *Mutation Research*, 475, 89–111. doi:10.1016/S0027-5107(01)00073-2.
- Fiala, M., Cribbs, D. H., Rosenthal, M., & Bernard, G. (2007). Phagocytosis of amyloid-beta and inflammation: Two faces of innate immunity in Alzheimer's disease. *Journal of Alzheimer's Disease*, 11, 457–463.
- Frautschy, S. A., Hu, W., Kim, P., Miller, S. A., Chu, T., Harris-White, M. E., et al. (2001). Phenolic anti-inflammatory antioxidant reversal of Abeta-induced cognitive deficits and neuropathology. *Neurobiology of Aging*, 22, 993–1005. doi:10.1016/S0197-4580(01)00300-1.
- Gao, D., Zhang, X., Jiang, X., Peng, Y., Huang, W., Cheng, G., et al. (2006a). Resveratrol reduces the elevated level of MMP-9 induced by cerebral ischemia-reperfusion in mice. *Life Sciences*, 78, 2564–2570. doi:10.1016/j.lfs.2005.10.030.
- Gao, H. M., Hong, J. S., Zhang, W., & Liu, B. (2002). Distinct role for microglia in rotenone-induced degeneration of dopaminergic neurons. *Journal of Neuroscience*, 22, 782–790.
- Gao, H. M., Hong, J. S., Zhang, W., & Liu, B. (2003a). Synergistic dopaminergic neurotoxicity of the pesticide rotenone and inflammogen lipopolysaccharide: Relevance to the etiology of Parkinson's disease. *Journal of Neuroscience*, 23, 1228–1236.
- Gao, H. M., Liu, B., Zhang, W., & Hong, J. S. (2003b). Critical role of microglial NADPH oxidase-derived free radicals in the in vitro MPTP model of Parkinson's disease. *FASEB Journal*, 17, 1954–1956.
- Gao, H. M., Liu, B., Zhang, W., & Hong, J. S. (2003c). Synergistic dopaminergic neurotoxicity of MPTP and inflammogen lipopolysaccharide: Relevance to the etiology of Parkinson's disease. *FASEB Journal*, 17, 1957–1959.
- Gao, Z. B., Chen, X. Q., & Hu, G. Y. (2006b). Inhibition of excitatory synaptic transmission by trans-resveratrol in rat hippocampus. *Brain Research*, 1111, 41–47. doi:10.1016/j.brainres.2006.06.096.
- Garcia-Alloza, M., Dodwell, S. A., Meyer-Luehmann, M., Hyman, B. T., & Bacskai, B. J. (2006). Plaque-derived oxidative stress mediates distorted neurite trajectories in the Alzheimer mouse model. *Journal of Neuropathology and Experimental Neurology*, 65, 1082–1089. doi:10.1097/01.jnen.0000240468.12543.af.
- Gelinas, S., & Martinoli, M. G. (2002). Neuroprotective effect of estradiol and phytoestrogens on MPP+-induced cytotoxicity in neuronal PC12 cells. *Journal of Neuroscience Research*, 70, 90–96. doi:10.1002/jnr.10315.
- Goel, A., Kunnumakara, A. B., & Aggarwal, B. B. (2008). Curcumin as “Curecumin”: From kitchen to clinic. *Biochemical Pharmacology*, 75, 787–809. doi:10.1016/j.bcp.2007.08.016.
- Gonzalez-Hernandez, T., Perez de la Cruz, M. A., & Mantolan-Sarmiento, B. (1996). Histochemical and immunohistochemical detection of neurons that produce nitric oxide: Effect of different fixative parameters and immunoreactivity against non-neuronal NOS antisera. *Journal of Histochemistry and Cytochemistry*, 44, 1399–1413.
- Guo, S., Yan, J., Yang, T., Yang, X., Bezdard, E., & Zhao, B. (2007). Protective effects of green tea polyphenols in the 6-OHDA rat model of Parkinson's disease through inhibition of ROS-NO pathway. *Biological Psychiatry*, 62, 1353–1362. doi:10.1016/j.biopsych.2007.04.020.
- Halliwel, B. (2006). Oxidative stress and neurodegeneration: Where are we now? *Journal of Neurochemistry*, 97, 1634–1658. doi:10.1111/j.1471-4159.2006.03907.x.
- Halliwel, B. (2008). Are polyphenols antioxidants or pro-oxidants? What do we learn from cell culture and in vivo studies? *Archives of Biochemistry and Biophysics*, 476, 107–112. doi:10.1016/j.abb.2008.01.028.
- Han, Y. S., Zheng, W. H., Bastianetto, S., Chabot, J. G., & Quirion, R. (2004). Neuroprotective effects of resveratrol against beta-amyloid-induced neurotoxicity in rat hippocampal neurons: Involvement of protein kinase C. *British Journal of Pharmacology*, 141, 997–1005. doi:10.1038/sj.bjp.0705688.
- Hartman, R. E., Shah, A., Fagan, A. M., Schwetye, K. E., Parsadanian, M., Schulman, R. N., et al. (2006). Pomegranate juice decreases amyloid load and improves behavior in a mouse model of Alzheimer's disease. *Neurobiology of Disease*, 24, 506–515. doi:10.1016/j.nbd.2006.08.006.
- Hong, J. T., Ryu, S. R., Kim, H. J., et al. (2000). Neuroprotective effect of green tea extract in experimental ischemia-reperfusion brain injury. *Brain Research Bulletin*, 53, 743–749. doi:10.1016/S0361-9230(00)00348-8.
- Hong, J. T., Ryu, S. R., Kim, H. J., Lee, J. K., Lee, S. H., Yun, Y. P., et al. (2001). Protective effect of green tea extract on ischemia/reperfusion-induced brain injury in Mongolian gerbils. *Brain Research*, 888, 11–18. doi:10.1016/S0006-8993(00)02935-8.
- Howitz, K. T., Bitterman, K. J., Cohen, H. Y., et al. (2003). Small molecule activators of sirtuins extend *Saccharomyces cerevisiae* lifespan. *Nature*, 425, 191–196. doi:10.1038/nature01960.
- Infanger, D. W., Sharma, R. V., & Davisson, R. L. (2006). NADPH oxidases of the brain: Distribution, regulation, and function. *Antioxidants & Redox Signaling*, 8, 1583–1596. doi:10.1089/ars.2006.8.1583.
- Jagatha, B., Mythri, R. B., Vali, S., & Bharath, M. M. (2008). Curcumin treatment alleviates the effects of glutathione depletion in vitro and in vivo: Therapeutic implications for Parkinson's disease explained via in silico studies. *Free Radical Biology and Medicine*, 44, 907–917. doi:10.1016/j.freeradbiomed.2007.11.011.
- Jang, J. H., & Surh, Y. J. (2003). Protective effect of resveratrol on beta-amyloid-induced oxidative PC12 cell death. *Free Radical Biology and Medicine*, 34, 1100–1110. doi:10.1016/S0891-5849(03)00062-5.
- Jekabsone, A., Mander, P. K., Tickler, A., Sharpe, M., & Brown, G. C. (2006). Fibrillar beta-amyloid peptide Abeta1–40 activates microglial proliferation via stimulating TNF-alpha release and H2O2 derived from NADPH oxidase: A cell culture study. *Journal of Neuroinflammation*, 3, 24. doi:10.1186/1742-2094-3-24.
- Jiang, J., Wang, W., Sun, Y. J., Hu, M., Li, F., & Zhu, D. Y. (2007). Neuroprotective effect of curcumin on focal cerebral ischemic rats by preventing blood-brain barrier damage. *European Journal of Pharmacology*, 561, 54–62. doi:10.1016/j.ejphar.2006.12.028.
- Joseph, J. A., Denisova, N. A., Arendash, G., Gordon, M., Diamond, D., Shukitt-Hale, B., et al. (2003). Blueberry supplementation enhances signaling and prevents behavioral deficits in an Alzheimer disease model. *Nutritional Neuroscience*, 6, 153–162. doi:10.1080/102841503100011282.
- Kahles, T., Luedike, P., Endres, M., Galla, H. J., Steinmetz, H., Busse, R., et al. (2007). NADPH oxidase plays a central role in blood-brain barrier damage in experimental stroke. *Stroke*, 38, 3000–3006. doi:10.1161/STROKEAHA.107.489765.
- Kang, T. H., Hur, J. Y., Kim, H. B., Ryu, J. H., & Kim, S. Y. (2006). Neuroprotective effects of the cyanidin-3-O-beta-D-glucopyranoside isolated from mulberry fruit against cerebral ischemia. *Neuroscience Letters*, 391, 122–126. doi:10.1016/j.neulet.2005.08.053.
- Kim, M. S., Lee, J. I., Lee, W. Y., & Kim, S. E. (2004). Neuroprotective effect of *Ginkgo biloba* L. extract in a rat model of Parkinson's disease. *Phytotherapy Research*, 18, 663–666. doi:10.1002/ptr.1486.
- Kim, Y. A., Kim, G. Y., Park, K. Y., & Choi, Y. H. (2007). Resveratrol inhibits nitric oxide and prostaglandin E2 production

- by lipopolysaccharide-activated C6 microglia. *Journal of Medicinal Food*, 10, 218–224. doi:10.1089/jmf.2006.143.
- Kim, Y. A., Lim, S. Y., Rhee, S. H., Park, K. Y., Kim, C. H., Choi, B. T., et al. (2006). Resveratrol inhibits inducible nitric oxide synthase and cyclooxygenase-2 expression in beta-amyloid-treated C6 glioma cells. *International Journal of Molecular Medicine*, 17, 1069–1075.
- Kishida, K. T., & Klann, E. (2007). Sources and targets of reactive oxygen species in synaptic plasticity and memory. *Antioxidants & Redox Signaling*, 9, 233–244. doi:10.1089/ars.2007.9.233.
- Kishida, K. T., Pao, M., Holland, S. M., & Klann, E. (2005). NADPH oxidase is required for NMDA receptor-dependent activation of ERK in hippocampal area CA1. *Journal of Neurochemistry*, 94, 299–306. doi:10.1111/j.1471-4159.2005.03189.x.
- Koshimura, I., Imai, H., Hidano, T., Endo, K., Mochizuki, H., Kondo, T., et al. (1997). Dimethoxyphenylethylamine and tetrahydropapaverine are toxic to the nigrostriatal system. *Brain Research*, 773, 108–116. doi:10.1016/S0006-8993(97)00922-0.
- Kotilinek, L. A., Westerman, M. A., Wang, Q., et al. (2008). Cyclooxygenase-2 inhibition improves amyloid-beta-mediated suppression of memory and synaptic plasticity. *Brain*, 131, 651–664. doi:10.1093/brain/awn008.
- Kriem, B., Sponne, I., Fifre, A., et al. (2005). Cytosolic phospholipase A2 mediates neuronal apoptosis induced by soluble oligomers of the amyloid-beta peptide. *FASEB Journal*, 19, 85–87.
- Lambeth, J. D. (2007). Nox enzymes, ROS, and chronic disease: An example of antagonistic pleiotropy. *Free Radical Biology and Medicine*, 43, 332–347. doi:10.1016/j.freeradbiomed.2007.03.027.
- Langston, J. W., Irwin, I., & Ricaurte, G. A. (1987). Neurotoxins, parkinsonism and Parkinson's disease. *Pharmacology and Therapeutics*, 32, 19–49. doi:10.1016/0163-7258(87)90062-3.
- Levites, Y., Amit, T., Youdim, M. B., & Mandel, S. (2002a). Involvement of protein kinase C activation and cell survival/cell cycle genes in green tea polyphenol (-)-epigallocatechin 3-gallate neuroprotective action. *Journal of Biological Chemistry*, 277, 30574–30580. doi:10.1074/jbc.M202832200.
- Levites, Y., Youdim, M. B., Maor, G., & Mandel, S. (2002b). Attenuation of 6-hydroxydopamine (6-OHDA)-induced nuclear factor-kappaB (NF-kappaB) activation and cell death by tea extracts in neuronal cultures. *Biochemical Pharmacology*, 63, 21–29. doi:10.1016/S0006-2952(01)00813-9.
- Li, M., Pisalyaput, K., Galvan, M., & Tenner, A. J. (2004). Macrophage colony stimulatory factor and interferon-gamma trigger distinct mechanisms for augmentation of beta-amyloid-induced microglia-mediated neurotoxicity. *Journal of Neurochemistry*, 91, 623–633. doi:10.1111/j.1471-4159.2004.02765.x.
- Li, X., & Sun, A. Y. (1999). Paraquat induced activation of transcription factor AP-1 and apoptosis in PC12 cells. *Journal of Neural Transmission*, 106, 1–21. doi:10.1007/s007020050137.
- Lim, G. P., Chu, T., Yang, F., Beech, W., Frautschy, S. A., & Cole, G. M. (2001). The curry spice curcumin reduces oxidative damage and amyloid pathology in an Alzheimer transgenic mouse. *Journal of Neuroscience*, 21, 8370–8377.
- Lim, H. J., Lee, K. S., Lee, S., Park, J. H., Choi, H. E., Go, S. H., et al. (2007a). 15d-PGJ2 stimulates HO-1 expression through p38 MAP kinase and Nrf-2 pathway in rat vascular smooth muscle cells. *Toxicology and Applied Pharmacology*, 223, 20–27. doi:10.1016/j.taap.2007.04.019.
- Lim, M. L., Mercer, L. D., Nagley, P., & Beart, P. M. (2007b). Rotenone and MPP+ preferentially redistribute apoptosis-inducing factor in apoptotic dopamine neurons. *NeuroReport*, 18, 307–312. doi:10.1097/WNR.0b013e32801b3ca6.
- Liou, H. H., Tsai, M. C., Chen, C. J., Jeng, J. S., Chang, Y. C., Chen, S. Y., et al. (1997). Environmental risk factors and Parkinson's disease: A case-control study in Taiwan. *Neurology*, 48, 1583–1588.
- Lu, K. T., Chiou, R. Y., Chen, L. G., Chen, M. H., Tseng, W. T., Hsieh, H. T., et al. (2006). Neuroprotective effects of resveratrol on free radical ischemia-induced neuron loss mediated by free radical scavenging and cerebral blood flow elevation. *Journal of Agricultural and Food Chemistry*, 54, 3126–3131. doi:10.1021/jf053011q.
- Manach, C., & Donovan, J. L. (2004). Pharmacokinetics and metabolism of dietary flavonoids in humans. *Free Radical Research*, 38, 771–785. doi:10.1080/10715760410001727858.
- Mancuso, C., Scapagini, G., Curro, D., Giuffrida Stella, A. M., De Marco, C., Butterfield, D. A., et al. (2007). Mitochondrial dysfunction, free radical generation and cellular stress response in neurodegenerative disorders. *Frontiers in Bioscience*, 12, 1107–1123. doi:10.2741/2130.
- Mandel, S., Weinreb, O., Amit, T., & Youdim, M. B. (2004). Cell signaling pathways in the neuroprotective actions of the green tea polyphenol (-)-epigallocatechin-3-gallate: Implications for neurodegenerative diseases. *Journal of Neurochemistry*, 88, 1555–1569.
- Mandel, S., & Youdim, M. B. (2004). Catechin polyphenols: Neurodegeneration and neuroprotection in neurodegenerative diseases. *Free Radical Biology and Medicine*, 37, 304–317. doi:10.1016/j.freeradbiomed.2004.04.012.
- Mander, P. K., Jekabsone, A., & Brown, G. C. (2006). Microglia proliferation is regulated by hydrogen peroxide from NADPH oxidase. *Journal of Immunology*, 176, 1046–1052.
- Marambaud, P., Zhao, H., & Davies, P. (2005). Resveratrol promotes clearance of Alzheimer's disease amyloid-beta peptides. *Journal of Biological Chemistry*, 280, 37377–37382. doi:10.1074/jbc.M508246200.
- Masuda, M., Suzuki, N., Taniguchi, S., Oikawa, T., Nonaka, T., Iwatsubo, T., et al. (2006). Small molecule inhibitors of alpha-synuclein filament assembly. *Biochemistry*, 45, 6085–6094. doi:10.1021/bi0600749.
- Mattson, M. P. (2007). Calcium and neurodegeneration. *Aging Cell*, 6, 337–350. doi:10.1111/j.1474-9726.2007.00275.x.
- Mattson, M. P. (2008). Dietary factors, hormesis and health. *Ageing Research Reviews*, 7, 43–48. doi:10.1016/j.arr.2007.08.004.
- McGeer, P. L., Itagaki, S., Akiyama, H., & McGeer, E. G. (1988). Rate of cell death in parkinsonism indicates active neuropathological process. *Annals of Neurology*, 24, 574–576. doi:10.1002/ana.410240415.
- McKeel, D. W., Jr., Price, J. L., Miller, J. P., Grant, E. A., Xiong, C., Berg, L., et al. (2004). Neuropathologic criteria for diagnosing Alzheimer disease in persons with pure dementia of Alzheimer type. *Journal of Neuropathology and Experimental Neurology*, 63, 1028–1037.
- Mercer, L. D., Kelly, B. L., Horne, M. K., & Beart, P. M. (2005). Dietary polyphenols protect dopamine neurons from oxidative insults and apoptosis: Investigations in primary rat mesencephalic cultures. *Biochemical Pharmacology*, 69, 339–345. doi:10.1016/j.bcp.2004.09.018.
- Miller, R. L., James-Kracke, M., Sun, G. Y., & Sun, A. Y. (2008). Oxidative and inflammatory pathways in Parkinson's disease. *Neurochemical Research*. doi:10.1007/s11064-008-9656-2.
- Miller, R. L., Sun, G. Y., & Sun, A. Y. (2007). Cytotoxicity of paraquat in microglial cells: Involvement of PKCdelta- and ERK1/2-dependent NADPH oxidase. *Brain Research*, 1167, 129–139. doi:10.1016/j.brainres.2007.06.046.
- Morelli, V., & Naquin, C. (2002). Alternative therapies for traditional disease states: Menopause. *American Family Physician*, 66, 129–134.
- Ndiaye, M., Chataigneau, M., Lobysheva, I., Chataigneau, T., & Schini-Kerth, V. B. (2005). Red wine polyphenol-induced, endothelium-dependent NO-mediated relaxation is due to the redox-sensitive PI3-kinase/Akt-dependent phosphorylation of

- endothelial NO-synthase in the isolated porcine coronary artery. *FASEB Journal*, 19, 455–457.
- Ono, K., Hasegawa, K., Naiki, H., & Yamada, M. (2004). Curcumin has potent anti-amyloidogenic effects for Alzheimer's beta-amyloid fibrils in vitro. *Journal of Neuroscience Research*, 75, 742–750. doi:10.1002/jnr.20025.
- Ono, K., Naiki, H., & Yamada, M. (2006). The development of preventives and therapeutics for Alzheimer's disease that inhibit the formation of beta-amyloid fibrils (fA $\beta$ ), as well as destabilize preformed fA $\beta$ . *Current Pharmaceutical Design*, 12, 4357–4375. doi:10.2174/138161206778793010.
- Ono, K., & Yamada, M. (2006). Antioxidant compounds have potent anti-fibrillogenic and fibril-destabilizing effects for alpha-synuclein fibrils in vitro. *Journal of Neurochemistry*, 97, 105–115. doi:10.1111/j.1471-4159.2006.03707.x.
- Pandey, N., Strider, J., Nolan, W. C., Yan, S. X., & Galvin, J. E. (2008). Curcumin inhibits aggregation of alpha-synuclein. *Acta Neuropathologica*, 115, 479–489. doi:10.1007/s00401-007-0332-4.
- Pu, F., Mishima, K., Egashira, N., et al. (2004). Protective effect of buckwheat polyphenols against long-lasting impairment of spatial memory associated with hippocampal neuronal damage in rats subjected to repeated cerebral ischemia. *Journal of Pharmacological Sciences*, 94, 393–402. doi:10.1254/jphs.94.393.
- Rahman, M., Riaz, M., & Desai, U. R. (2007). Synthesis of biologically relevant biflavonoids—A review. *Chemistry and Biodiversity*, 4, 2495–2527. doi:10.1002/cbdv.200790205.
- Ramassamy, C. (2006). Emerging role of polyphenolic compounds in the treatment of neurodegenerative diseases: A review of their intracellular targets. *European Journal of Pharmacology*, 545, 51–64. doi:10.1016/j.ejphar.2006.06.025.
- Raskin, I., Ribnicky, D. M., Komarnytsky, S., et al. (2002). Plants and human health in the twenty-first century. *Trends in Biotechnology*, 20, 522–531. doi:10.1016/S0167-7799(02)02080-2.
- Rathore, P., Dohare, P., Varma, S., Ray, A., Sharma, U., Jaganathanan, N. R., et al. (2008). Curcuma oil: Reduces early accumulation of oxidative product and is anti-apoptogenic in transient focal ischemia in rat brain. *Neurochemical Research*, 33, 1672–1682.
- Raval, A. P., Dave, K. R., & Perez-Pinzon, M. A. (2006). Resveratrol mimics ischemic preconditioning in the brain. *Journal of Cerebral Blood Flow and Metabolism*, 26, 1141–1147.
- Rezaei-Zadeh, K., Shytle, D., Sun, N., et al. (2005). Green tea epigallocatechin-3-gallate (EGCG) modulates amyloid precursor protein cleavage and reduces cerebral amyloidosis in Alzheimer transgenic mice. *Journal of Neuroscience*, 25, 8807–8814. doi:10.1523/JNEUROSCI.1521-05.2005.
- Rice-Evans, C., & Miller, N. (1997). Measurement of the antioxidant status of dietary constituents, low density lipoproteins and plasma. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 57, 499–505. doi:10.1016/S0952-3278(97)90435-X.
- Ringman, J. M., Frautschy, S. A., Cole, G. M., Masterman, D. L., & Cummings, J. L. (2005). A potential role of the curry spice curcumin in Alzheimer's disease. *Current Alzheimer Research*, 2, 131–136. doi:10.2174/1567205053585882.
- Ritz, M. F., Ratajczak, P., Curin, Y., Cam, E., Mendelowitsch, A., Pinet, F., et al. (2008). Chronic treatment with red wine polyphenol compounds mediates neuroprotection in a rat model of ischemic cerebral stroke. *The Journal of Nutrition*, 138, 519–525.
- Ross, J. A., & Kasum, C. M. (2002). Dietary flavonoids: Bioavailability, metabolic effects, and safety. *Annual Review of Nutrition*, 22, 19–34. doi:10.1146/annurev.nutr.22.111401.144957.
- Saito, A., Maier, C. M., Narasimhan, P., et al. (2005). Oxidative stress and neuronal death/survival signaling in cerebral ischemia. *Molecular Neurobiology*, 31, 105–116. doi:10.1385/MN:31:1-3:105.
- Saleem, S., Ahmad, M., Ahmad, A. S., Yousuf, S., Ansari, M. A., Khan, M. B., et al. (2006). Behavioral and histologic neuroprotection of aqueous garlic extract after reversible focal cerebral ischemia. *Journal of Medicinal Food*, 9, 537–544. doi:10.1089/jmf.2006.9.537.
- Salvioli, S., Sikora, E., Cooper, E. L., & Franceschi, C. (2007). Curcumin in cell death processes: A challenge for CAM of age-related pathologies. *Evidence-Based Complementary and Alternative Medicine*, 4, 181–190. doi:10.1093/ecam/nem043.
- Sang, N., & Chen, C. (2006). Lipid signaling and synaptic plasticity. *Neuroscientist*, 12, 425–434. doi:10.1177/1073858406290794.
- Santangelo, C., Vari, R., Scazzocchio, B., Di Benedetto, R., Filesi, C., & Masella, R. (2007). Polyphenols, intracellular signalling and inflammation. *Annali dell'Istituto Superiore di Sanita*, 43, 394–405.
- Schapiro, A. H. (1996). Neurotoxicity and the mechanisms of cell death in Parkinson's disease. *Advances in Neurology*, 69, 161–165.
- Schober, A. (2004). Classic toxin-induced animal models of Parkinson's disease: 6-OHDA and MPTP. *Cell and Tissue Research*, 318, 215–224. doi:10.1007/s00441-004-0938-y.
- Schroeter, H., Bahia, P., Spencer, J. P., Sheppard, O., Rattray, M., Cadenas, E., et al. (2007). (-)Epicatechin stimulates ERK-dependent cyclic AMP response element activity and up-regulates GluR2 in cortical neurons. *Journal of Neurochemistry*, 101, 1596–1606. doi:10.1111/j.1471-4159.2006.04434.x.
- Selkoe, D. J. (2001). Alzheimer's disease results from the cerebral accumulation and cytotoxicity of amyloid beta-protein. *Journal of Alzheimer's Disease*, 3, 75–80.
- Sharma, M., & Gupta, Y. K. (2002). Chronic treatment with trans resveratrol prevents intracerebroventricular streptozotocin induced cognitive impairment and oxidative stress in rats. *Life Sciences*, 71, 2489–2498. doi:10.1016/S0024-3205(02)02083-0.
- Shelat, P. B., Chalimoniuk, M., Wang, J. H., Strosznajder, J. B., Lee, J. C., Sun, A. Y., et al. (2008). Amyloid beta peptide and NMDA induce ROS from NADPH oxidase and AA release from cytosolic phospholipase A(2) in cortical neurons. *Journal of Neurochemistry*, 106, 45–55.
- Shen, L., & Zhang, J. (2003). Ginsenoside Rg1 increases ischemia-induced cell proliferation and survival in the dentate gyrus of adult gerbils. *Neuroscience Letters*, 344, 1–4. doi:10.1016/S0304-3940(03)00318-5.
- Shibata, H., Katsuki, H., Okawara, M., Kume, T., & Akaike, A. (2006). c-Jun N-terminal kinase inhibition and alpha-tocopherol protect midbrain dopaminergic neurons from interferon-gamma/lipopolysaccharide-induced injury without affecting nitric oxide production. *Journal of Neuroscience Research*, 83, 102–109. doi:10.1002/jnr.20700.
- Shin, D. H., Bae, Y. C., Kim-Han, J. S., Lee, J. H., Choi, I. Y., Son, K. H., et al. (2006). Polyphenol amentoflavone affords neuroprotection against neonatal hypoxic-ischemic brain damage via multiple mechanisms. *Journal of Neurochemistry*, 96, 561–572. doi:10.1111/j.1471-4159.2005.03582.x.
- Simonian, N. A., & Coyle, J. T. (1996). Oxidative stress in neurodegenerative diseases. *Annual Review of Pharmacology and Toxicology*, 36, 83–106. doi:10.1146/annurev.pa.36.040196.000503.
- Simonyi, A., Wang, Q., Miller, R. L., Yusof, M., Shelat, P. B., Sun, A. Y., et al. (2005). Polyphenols in cerebral ischemia: Novel targets for neuroprotection. *Molecular Neurobiology*, 31, 135–147. doi:10.1385/MN:31:1-3:135.
- Simonyi, A., Woods, D., Sun, A. Y., & Sun, G. Y. (2002). Grape polyphenols inhibit chronic ethanol-induced COX-2 mRNA expression in rat brain. *Alcoholism, Clinical and Experimental Research*, 26, 352–357.
- Snyder, E. M., Nong, Y., Almeida, C. G., et al. (2005). Regulation of NMDA receptor trafficking by amyloid-beta. *Nature Neuroscience*, 8, 1051–1058. doi:10.1038/nn1503.

- Stackman, R. W., Eckenstein, F., Frei, B., Kulhanek, D., Nowlin, J., & Quinn, J. F. (2003). Prevention of age-related spatial memory deficits in a transgenic mouse model of Alzheimer's disease by chronic *Ginkgo biloba* treatment. *Experimental Neurology*, *184*, 510–520. doi:10.1016/S0014-4886(03)00399-6.
- Stolk, J., Hiltermann, T. J., Dijkman, J. H., & Verhoeven, A. J. (1994). Characteristics of the inhibition of NADPH oxidase activation in neutrophils by apocynin, a methoxy-substituted catechol. *American Journal of Respiratory Cell and Molecular Biology*, *11*, 95–102.
- Storch, A., Jost, W. H., Vieregge, P., et al. (2007). Randomized, double-blind, placebo-controlled trial on symptomatic effects of coenzyme Q(10) in Parkinson disease. *Archives of Neurology*, *64*, 938–944. doi:10.1001/archneur.64.7.nct60005.
- Sun, A. Y., & Chen, Y. M. (1998). Oxidative stress and neurodegenerative disorders. *Journal of Biomedical Science*, *5*, 401–414. doi:10.1007/BF02255928.
- Sun, A. Y., Yang, W. L., & Kim, H. D. (1993). Free radical and lipid peroxidation in manganese-induced neuronal cell injury. *Annals of the New York Academy of Sciences*, *679*, 358–363. doi:10.1111/j.1749-6632.1993.tb18322.x.
- Sun, G. Y., Xia, J., Draczynska-Lusiak, B., Simonyi, A., & Sun, A. Y. (1999a). Grape polyphenols protect neurodegenerative changes induced by chronic ethanol administration. *NeuroReport*, *10*, 93–96. doi:10.1097/00001756-199901180-00018.
- Sun, G. Y., Xia, J., Xu, J., Allenbrand, B., Simonyi, A., Rudeen, P. K., et al. (1999b). Dietary supplementation of grape polyphenols to rats ameliorates chronic ethanol-induced changes in hepatic morphology without altering changes in hepatic lipids. *The Journal of Nutrition*, *129*, 1814–1819.
- Sutherland, B. A., Rahman, R. M., & Appleton, I. (2006). Mechanisms of action of green tea catechins, with a focus on ischemia-induced neurodegeneration. *The Journal of Nutritional Biochemistry*, *17*, 291–306. doi:10.1016/j.jnutbio.2005.10.005.
- Sweeney, M. I., Kalt, W., MacKinnon, S. L., Ashby, J., & Gottschall-Pass, K. T. (2002). Feeding rats diets enriched in lowbush blueberries for six weeks decreases ischemia-induced brain damage. *Nutritional Neuroscience*, *5*, 427–431. doi:10.1080/1028415021000055970.
- Tang, L. L., Ye, K., Yang, X. F., & Zheng, J. S. (2007). Apocynin attenuates cerebral infarction after transient focal ischaemia in rats. *Journal of International Medical Research*, *35*, 517–522.
- Tchantchou, F., Xu, Y., Wu, Y., Christen, Y., & Luo, Y. (2007). EGB 761 enhances adult hippocampal neurogenesis and phosphorylation of CREB in transgenic mouse model of Alzheimer's disease. *FASEB Journal*, *21*, 2400–2408. doi:10.1096/fj.06-7649com.
- Tohda, C., Matsumoto, N., Zou, K., Meselhy, M. R., & Komatsu, K. (2004). Abeta(25–35)-induced memory impairment, axonal atrophy, and synaptic loss are ameliorated by M1, A metabolite of protopanaxadiol-type saponins. *Neuropsychopharmacology*, *29*, 860–868. doi:10.1038/sj.npp.1300388.
- Tsai, S. K., Hung, L. M., Fu, Y. T., Cheng, H., Nien, M. W., Liu, H. Y., et al. (2007). Resveratrol neuroprotective effects during focal cerebral ischemia injury via nitric oxide mechanism in rats. *Journal of Vascular Surgery*, *46*, 346–353. doi:10.1016/j.jvs.2007.04.044.
- Vafeiadou, K., Vauzour, D., & Spencer, J. P. (2007). Neuroinflammation and its modulation by flavonoids. *Endocrine, Metabolic & Immune Disorders Drug Targets*, *7*, 211–224. doi:10.2174/187153007781662521.
- Vauzour, D., Vafeiadou, K., Corona, G., Pollard, S. E., Tzounis, X., & Spencer, J. P. (2007). Champagne wine polyphenols protect primary cortical neurons against peroxynitrite-induced injury. *Journal of Agricultural and Food Chemistry*, *55*, 2854–2860. doi:10.1021/jf063304z.
- Voko, Z., Hollander, M., Hofman, A., Koudstaal, P. J., & Breteler, M. M. (2003). Dietary antioxidants and the risk of ischemic stroke: The Rotterdam Study. *Neurology*, *61*, 1273–1275.
- Wang, C. N., Chi, C. W., Lin, Y. L., Chen, C. F., & Shiao, Y. J. (2001). The neuroprotective effects of phytoestrogens on amyloid beta protein-induced toxicity are mediated by abrogating the activation of caspase cascade in rat cortical neurons. *Journal of Biological Chemistry*, *276*, 5287–5295. doi:10.1074/jbc.M006406200.
- Wang, J., Ho, L., Zhao, Z., et al. (2006a). Moderate consumption of Cabernet Sauvignon attenuates Abeta neuropathology in a mouse model of Alzheimer's disease. *FASEB Journal*, *20*, 2313–2320. doi:10.1096/fj.06-6281com.
- Wang, Q., Simonyi, A., Li, W., Sisk, B. A., Miller, R. L., Macdonald, R. S., et al. (2005a). Dietary grape supplement ameliorates cerebral ischemia-induced neuronal death in gerbils. *Molecular Nutrition & Food Research*, *49*, 443–451. doi:10.1002/mnfr.200500019.
- Wang, Q., Smith, R. E., Luchtefeld, R., Sun, A. Y., Simonyi, A., Luo, R., et al. (2008). Bioavailability of apocynin through its conversion to glycoconjugate but not to diapocynin. *Phytomedicine*, *15*, 496–503.
- Wang, Q., Sun, A. Y., Simonyi, A., et al. (2005b). Neuroprotective mechanisms of curcumin against cerebral ischemia-induced neuronal apoptosis and behavioral deficits. *Journal of Neuroscience Research*, *82*, 138–148. doi:10.1002/jnr.20610.
- Wang, Q., Tompkins, K. D., Simonyi, A., Korhuis, R. J., Sun, A. Y., & Sun, G. Y. (2006b). Apocynin protects against global cerebral ischemia-reperfusion-induced oxidative stress and injury in the gerbil hippocampus. *Brain Research*, *1090*, 182–189. doi:10.1016/j.brainres.2006.03.060.
- Wang, Q., Xu, J., Rottinghaus, G. E., Simonyi, A., Lubahn, D., Sun, G. Y., et al. (2002). Resveratrol protects against global cerebral ischemic injury in gerbils. *Brain Research*, *958*, 439–447. doi:10.1016/S0006-8993(02)03543-6.
- Wang, Q., Yu, S., Simonyi, A., Sun, G. Y., & Sun, A. Y. (2005c). Kainic acid-mediated excitotoxicity as a model for neurodegeneration. *Molecular Neurobiology*, *31*, 3–16. doi:10.1385/MN:31:1-3:003.
- Wang, X., Su, B., Perry, G., Smith, M. A., & Zhu, X. (2007). Insights into amyloid-beta-induced mitochondrial dysfunction in Alzheimer disease. *Free Radical Biology and Medicine*, *43*, 1569–1573. doi:10.1016/j.freeradbiomed.2007.09.007.
- Wang, Y., Chang, C. F., Chou, J., Chen, H. L., Deng, X., Harvey, B. K., et al. (2005d). Dietary supplementation with blueberries, spinach, or spirulina reduces ischemic brain damage. *Experimental Neurology*, *193*, 75–84. doi:10.1016/j.expneurol.2004.12.014.
- Weinreb, O., Amit, T., & Youdim, M. B. (2008). The application of proteomics for studying the neurorescue activity of the polyphenol (-)-epigallocatechin-3-gallate. *Archives of Biochemistry and Biophysics*, *476*, 152–160. doi:10.1016/j.abb.2008.01.004.
- Weinreb, O., Mandel, S., Amit, T., & Youdim, M. B. (2004). Neurological mechanisms of green tea polyphenols in Alzheimer's and Parkinson's diseases. *Journal of Nutritional Biochemistry*, *15*, 506–516. doi:10.1016/j.jnutbio.2004.05.002.
- West, T., Atzeva, M., & Holtzman, D. M. (2007). Pomegranate polyphenols and resveratrol protect the neonatal brain against hypoxic-ischemic injury. *Developmental Neuroscience*, *29*, 363–372. doi:10.1159/000105477.
- Wu, A., Ying, Z., & Gomez-Pinilla, F. (2006). Dietary curcumin counteracts the outcome of traumatic brain injury on oxidative stress, synaptic plasticity, and cognition. *Experimental Neurology*, *197*, 309–317. doi:10.1016/j.expneurol.2005.09.004.
- Wu, D. C., Teismann, P., Tieu, K., Vila, M., Jackson-Lewis, V., Ischiropoulos, H., et al. (2003). NADPH oxidase mediates

- oxidative stress in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine model of Parkinson's disease. *Proceedings of the National Academy of Sciences of the United States of America*, 100, 6145–6150. doi:10.1073/pnas.0937239100.
- Wu, X. F., Block, M. L., Zhang, W., Qin, L., Wilson, B., Zhang, W. Q., et al. (2005). The role of microglia in paraquat-induced dopaminergic neurotoxicity. *Antioxidants & Redox Signaling*, 7, 654–661. doi:10.1089/ars.2005.7.654.
- Yang, F., Lim, G. P., Begum, A. N., et al. (2005). Curcumin inhibits formation of amyloid beta oligomers and fibrils, binds plaques, and reduces amyloid in vivo. *Journal of Biological Chemistry*, 280, 5892–5901. doi:10.1074/jbc.M404751200.
- Yang, W., & Sun, A. Y. (1998a). Paraquat-induced free radical reaction in mouse brain microsomes. *Neurochemical Research*, 23, 47–53. doi:10.1023/A:1022497319548.
- Yang, W. L., & Sun, A. Y. (1998b). Paraquat-induced cell death in PC12 cells. *Neurochemical Research*, 23, 1387–1394. doi:10.1023/A:1020750706762.
- Yi, H., Akao, Y., Maruyama, W., Chen, K., Shih, J., & Naoi, M. (2006). Type A monoamine oxidase is the target of an endogenous dopaminergic neurotoxin, N-methyl(R)salsolinol, leading to apoptosis in SH-SY5Y cells. *Journal of Neurochemistry*, 96, 541–549. doi:10.1111/j.1471-4159.2005.03573.x.
- Youdim, K. A., & Joseph, J. A. (2001). A possible emerging role of phytochemicals in improving age-related neurological dysfunctions: A multiplicity of effects. *Free Radical Biology and Medicine*, 30, 583–594. doi:10.1016/S0891-5849(00)00510-4.
- Zbarsky, V., Datla, K. P., Parkar, S., Rai, D. K., Aruoma, O. I., & Dexter, D. T. (2005). Neuroprotective properties of the natural phenolic antioxidants curcumin and naringenin but not quercetin and fisetin in a 6-OHDA model of Parkinson's disease. *Free Radical Research*, 39, 1119–1125. doi:10.1080/10715760500233113.
- Zhang, W., Wang, T., Qin, L., Gao, H. M., Wilson, B., Ali, S. F., et al. (2004). Neuroprotective effect of dextromethorphan in the MPTP Parkinson's disease model: Role of NADPH oxidase. *FASEB Journal*, 18, 589–591.
- Zhu, D., Lai, Y., Shelat, P. B., Hu, C., Sun, G. Y., & Lee, J. C. (2006). Phospholipases A2 mediate amyloid-beta peptide-induced mitochondrial dysfunction. *Journal of Neuroscience*, 26, 11111–11119. doi:10.1523/JNEUROSCI.3505-06.2006.